

0006-2952(93)E0064-E

SHORT COMMUNICATIONS

Bestatin transport in rabbit intestinal brush-border membrane vesicles

(Received 9 August 1993; accepted 16 November 1993)

Abstract—The effect of papain treatment on bestatin uptake by rabbit intestinal brush-border membrane vesicles (BBMVs) was studied. Papain treatment of BBMVs effectively diminished aminopeptidase activity but not bestatin uptake by a H^+ /dipeptide cotransporter. Bestatin uptake by BBMVs was composed of two saturable components, and after papain treatment the high-affinity component disappeared while the low-affinity component was retained. These findings suggest that high- and low-affinity components represent bestatin binding to aminopeptidase and the true uptake by the H^+ /dipeptide cotransporter, respectively.

Key words: bestatin; dipeptide; intestinal absorption; brush-border membrane; aminopeptidase; papain treatment

Bestatin is an immunostimulant given orally with anticancer chemotherapy [1, 2]. It is a dipeptide containing a β -amino acid, and is a potent inhibitor of several aminopeptidases [3–5]. In intestinal BBMVs,* dipeptides and oral cephalosporins are transported via a H⁺/dipeptide cotransporter and bestatin is also a substrate for this transporter [6–11]. However, in contrast to other substrates, bestatin binds firmly to aminopeptidases [3, 5]. We also observed that the activity of aminopeptidase in rabbit BBMVs was inhibited by bestatin with an IC₅₀ value of 5.0 μ M (mean of two experiments). Therefore the apparent uptake of bestatin by BBMVs may include this binding [10].

Aminopeptidase in BBMVs can be removed by papain treatment [12, 13]. In this report we examined the effect of papain treatment to clarify the nature of the uptake of bestatin by BBMVs.

Materials and Methods

Bestatin and [³H]bestatin (12.7 GBq/mmol) (Nippon Kayaku Co., Tokyo, Japan) were gifts. Papain (EC 3.4.22.2) 2× crystallized suspension was obtained from the Sigma Chemical Co. (St Louis, MO, U.S.A.).

BBMVs were isolated from the small intestine of male rabbits by a calcium precipitation method [8]. Papain treatment of BBMVs was carried out as described previously [14]. The uptake of bestatin by BBMVs was measured by a rapid filtration technique [10].

The radioactivity of [3 H]bestatin was determined by liquid scintillation counting, using an external standard to correct for quench. Protein was determined by the Bio-Rad Protein Assay Kit with bovine γ -globulin as the standard [15].

Results and Discussion

The effect of papain treatment of BBMVs on aminopeptidase activity was examined. At a papain concentration of $0.20\,\mathrm{U/mg}$ BBMV protein, aminopeptidase activity was diminished to less than 10% of control (untreated BBMVs, 1.88 ± 0.19 ; papain-treated BBMVs, $0.16\pm0.08\,\mu\mathrm{mol/mg}$ protein/min, mean \pm SE of

eight experiments). We reported previously that bestatin uptake by rabbit intestinal BBMVs was actively driven by an inward H⁺ gradient via the H⁺/dipeptide cotransporter [10]. Figure 1A shows the effect of papain treatment on bestatin uptake by BBMVs. The uptake of bestatin was actively driven in the presence of an inward H⁺ gradient in both papain-treated and untreated BBMVs. The bestatin uptake per milligram protein was markedly increased by papain treatment. Similar results were obtained for the uptake of cephradine, an aminocephalosporin which is also transported by the H⁺/dipeptide cotransporter (data not

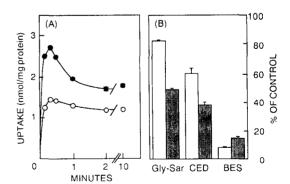


Fig. 1. Effect of papain treatment on bestatin uptake by BBMVs. BBMVs ($20 \,\mu\text{L}$, $175-192 \,\mu\text{g}$ protein for untreated BBMVs and $82-85 \,\mu\text{g}$ protein for papain-treated BBMVs), suspended in $100 \,\text{mM}$ mannitol, $100 \,\text{mM}$ KCl and $10 \,\text{mM}$ HEPES (pH 7.5), were incubated at 37° with the substrate mixture ($200 \,\mu\text{L}$) comprising $100 \,\text{mM}$ mannitol, $100 \,\text{mM}$ KCl, $10 \,\text{mM}$ Mes (pH 6.0) and $0.22 \,\text{mM}$ [^3H]bestatin. (A) Time course of bestatin uptake by untreated (\bigcirc) or papain-treated (\bigcirc) BBMVs. Each point represents the mean $^\pm$ SE of three determinations. (B) The uptake of [^3H]bestatin for $10 \,\text{sec}$ was measured in the absence (control) or presence of $10 \,\text{mM}$ glycylsarcosine (Gly-Sar), cephradine (CED) or unlabeled bestatin (BES). Untreated BBMVs (open column); papain-treated BBMVs (dotted column). Each column represents the mean $^\pm$ SE of three determinations.

^{*} Abbreviations: BBMVs, brush-border membrane vesicles; Mes, 2-(N-morpholino)ethanesulfonic acid.

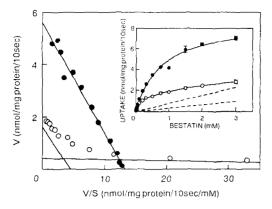


Fig. 2. Eadie-Hofstee plot of bestatin uptake by papaintreated or untreated BBMVs. BBMVs (20 µL, 245 µg protein for untreated BBMVs and 85 µg protein for papaintreated BBMVs), suspended in 100 mM mannitol, 100 mM KCl and 10 mM HEPES (pH 7.5), were incubated at 37° for 10 sec with the substrate mixture (200 µL) comprising 100 mM mannitol, 100 mM KCl, 10 mM Mes (pH 6.0) and varying concentrations of [3H]bestatin. The data were plotted after correction for the non-saturable component. The inset shows concentration-dependent curves of bestatin uptake and the broken lines represent the non-saturable components. Untreated BBMVs (O); papain-treated BBMVs (•). Each point represents the mean ± SE of three determinations. The kinetic parameters were calculated as described previously [10], and the values of estimates \pm SE were: $K_{m1} = 4.8 \pm 5.2 \,\mu\text{M}$, $V_{\text{max}1} =$ $2.48 \pm 0.77 \text{ nmol/mg protein/min}, K_{m2} = 0.36 \pm 0.13 \text{ mM},$ $V_{\text{max}2} = 9.67 \pm 0.96 \text{ nmol/mg} \text{ protein/min}, \ K_d = 1.89 \pm$ 0.35 nmol/mg protein/min/mM in untreated BBMVs, and $K_{\rm m} = 0.42 \pm 0.10 \, {\rm mM}$, $V_{\rm max} = 33.6 \pm 4.8 \, {\rm nmol/mg}$ protein/min, $K_{\rm d} = 4.54 \pm 1.36 \, {\rm nmol/mg}$ protein/min/mM in papain-treated BBMVs.

shown). Inhibition of bestatin uptake by the substrate for the $H^+/\text{dipeptide}$ cotransporter is shown in Fig. 1B. Glycylsarcosine, cephradine and unlabeled bestatin inhibited ${}^{1}\text{H}$]bestatin uptake in both papain-treated and untreated BBMVs, indicating that bestatin is taken up by a $H^+/\text{dipeptide}$ cotransport system in these BBMVs. The apparent uptake by untreated BBMVs should include a significant amount of bestatin binding to aminopeptidase, because the inhibitory effect of glycylsarcosine and cephradine, which do not bind to the enzyme as strongly as bestatin, was weaker in untreated than in papain-treated BBMVs. Thus, as described previously [12–14], papain treatment removed aminopeptidase and other membrane proteins without impairing bestatin transport by the $H^+/$ dipeptide cotransporter.

Figure 2 shows the concentration dependence of bestatin uptake by BBMVs. The uptake was saturable in both papain-treated and untreated BBMVs. As described previously [10], bestatin uptake by untreated BBMVs was composed of two saturable components. After papain treatment, the high-affinity component disappeared, while the low-affinity component was retained. Taken together,

these results indicate that the high- and low-affinity components of the uptake of bestatin by untreated BBMVs represent the binding to aminopeptidase and the true uptake by the H⁺/dipeptide cotransporter, respectively.

Department of Pharmacy Kyoto University Hospital Faculty of Medicine Kyoto University Sakyo-ku Kyoto 606-01, Japan

Mikihisa Takano* Yoshiko Tomita Toshiya Katsura Masato Yasuhara Ken-ichi Inui† Ryohei Hori‡

REFERENCES

- Ishizuka M, Masuda T, Kanbayashi N, Fukasawa S, Takeuchi T, Aoyagi T and Umezawa H, Effect of bestatin on mouse immune system and experimental murine tumors. J Antibiot 33: 642-652, 1980.
- Ota K, Review of ubenimex (Bestatin); clinical research. Biomed Pharmacother 45: 55-60, 1991.
- Umezawa H, Aoyagi T, Suda H, Hamada M and Takeuchi T, Bestatin, an inhibitor of aminopeptidase B, produced by actinomycetes. J Antibiot 29: 97–99, 1976
- 4. Suda H, Takita T, Aoyagi T and Umezawa H. The structure of bestatin. *J Antibiot* 29: 100–101, 1976.
- Wilkes SH and Prescott M, The slow, tight binding of bestatin and amastatin to aminopeptidases. J Biol Chem 260: 13154–13162, 1985.
- Ganapathy V, Burckhardt G and Leibach FH, Characteristics of glycylsarcosine transport in rabbit intestinal brush-border membrane vesicles. *J Biol Chem* 259: 8954–8959, 1984.
- 7. Hoshi T, Proton-coupled transport of organic solutes in animal cell membranes and its relation to Natransport. *Jpn J Physiol* **35**: 179–191, 1985.
- Okano T, Inui K, Maegawa H, Takano M and Hori R, H⁻ coupled uphill transport of aminocephalosporins via the dipeptide transport system in rabbit intestinal brush-border membranes. J Biol Chem 261: 14130– 14134, 1986.
- Tomita Y, Katsura T, Okano T, Inui K and Hori R, Transport mechanisms of bestatin in rabbit intestinal brush-border membranes; role of H*/dipeptide cotransport system. J Pharmacol Exp Ther 252: 859–862, 1990.
- Inui K, Tomita Y, Katsura T, Okano T, Takano M and Hori R, H⁻ coupled active transport of bestatin via the dipeptide transport system in rabbit intestinal brush-border membranes. *J Pharmacol Exp Ther* 260: 482-486, 1991.
- Kramer W, Dechent C, Girbig F, Gutjahr U and Neubauer H, Intestinal uptake of dipeptides and βlactam antibiotics. I. The intestinal uptake system for dipeptides and β-lactam antibiotics is not part of a brush border membrane peptidase. Biochim Biophys Acta 1030: 41-49, 1990.
- 12. Desnulle P, Intestinal and renal aminopeptidase: a model of a transmembrane protein. Eur J Biochem 101: 1–11, 1979.
- Ganapathy V, Mendicino JF and Leibach FH, Effect of papain treatment on dipeptide transport into rabbit intestinal brush border vesicles. *Life Sci* 29: 2451–2457, 1981.
- 14. Inui K, Okano T, Takano M, Saito H and Hori R, Carrier-mediated transport of cephalexin via the dipeptide transport system in rat renal brush-border membrane vesicles. *Biochim Biophys Acta* 769: 449– 454, 1984.
- 15. Bradford MM, A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248–254, 1976.

^{*} Corresponding author. Tel. (81) 75-751-3586; FAX (81) 75-751-3205.

[†] Present address: Department of Hospital Pharmacy, School of Medicine, Tokyo Medical and Dental University, Bunkyo-ku, Tokyo 113, Japan.

[‡] Present address: Pharmaceutical Research and Technology Institute, Kinki University, 3-4-1 Kowakae, Higashi-osaka 577, Japan.